

<https://doi.org/10.51470/IJNS.2024.01.01.1>

International Journal of Nature Science (IJNS)



From Forest to First Aid: A Review of Western Ghats Plants for Wound Healing

PURUSHOTHAMAN T^{1&2}, SARANYA N², ARUN KUMAR D¹

Department of Biotechnology

¹SNMV College of Arts and Science, Coimbatore, (TN) - India

² Nehru Arts & Science College, Coimbatore, (TN) - India

*Corresponding Author: purushonlines@gmail.com

Article History

Volume:1, Issue:1, 2024

Received: 17th January, 2024

Accepted: 20th January, 2024

Published: 30th January, 2024.

doi.org/10.51470/IJNS.2024.01.01.1-14

***Abstract:** India has been blessed with immense varieties of plants and most of them are found in the Western Ghats. There is a vast number of medicinal plants are found in the Western Ghats, which are widely used by the indigenous people. Medicinal plants have an essential role in healing and managing wounds. The bioactive compounds found in plants are responsible for its wound healing activities. The wound healing properties of the medicinal plants are influenced by the anti- microbial, anti-inflammatory, and anti-oxidant properties of plants. The selected medicinal plants possess remarkable inhibitory activity against the different bacterial as well as fungal species. Besides that, the results of the in-vitro studies have also reported the significant anti-inflammatory, anti-oxidant, and wound healing properties of those medicinal plants. Hence, the medicinal plants found in Western Ghats will be promising source for treating and healing the different types of wounds. The present review aims to document the scientific reports for the anti-microbial, anti- inflammatory, anti-oxidant and wound healing potency of some of the medicinal plants of Western Ghats.*

***Keywords:** Medicinal plants, Wound healing, Anti-microbial, Anti-inflammatory, Anti-oxidant*

Authors citation: Purushothaman.T et.al., From Forest to First Aid: A Review of Western Ghats Plants for Wound Healing.Int.J.Nat.Sci.Vol.1(1). 2024.Pp:1-10.

<https://doi.org/10.51470/IJNS.2024.01.01.1>

INTRODUCTION

The role of medicinal plants in human life is indispensable from the olden days, which are commonly used by indigenous people. But, 60-80% of worldwide population are still depends on the plant and derived components for treating diseases. India has been enriched with broad varieties of medicinal plants and most of them are found in Western Ghats (Dar, R. A et al., 2017). So, these medicinal plants are extensively used by the various tribal communities residing in and around Western Ghats (Jeyaprakash, K et al., 2011).

The medicinal plants play a pivotal role in treating and healing all types of wounds since ancient times. The bioactive components present in the plants forms the basis for the traditional as well as modern medicines. Majority of the available medicines are derived from plants, via the isolation of biologically active compounds. There are several scientific evidence are available in corresponding with healing of wounds using medicinal plants (Alam, G et al., 2011)

The plants will initiate the healing mechanism in a natural way and it is cost-effective solution than the synthetic medicines. Moreover, it exhibit lesser side-effects in comparison with chemical substances (Sharma, P et al., 2020). Considering these advantages, we have attempted to explore the wound healing activities of some medicinal plants found in the Western Ghats. Hence, this review provides the scientific evidence for the anti-microbial, anti-inflammatory, anti-oxidant and wound healing activity of selected medicinal plants of Western Ghats.

MECHANISM OF WOUND HEALING

The wound healing is the highly complex process which involves numerous events for the restoration or re-epithelialization of the impaired skin (Han, G., & Ceilley, R. 2017). It is a pivotal process, where the varieties of cells and its products synchronize to initiate the mechanism of wound healing. The duration of wound healing will differ depending on the types of wounds. The mechanism of wound healing begins immediately after the injury and it may continue for varied time period (Rawat, S et al., 2012). There are different phases involved in the wound healing mechanism such as

- Hemostatic phase
- Inflammatory phase
- Proliferative phase
- Remodeling phase

Hemostatic phase:

The mechanism of wound healing begins with the hemostatic phase, which happens immediately after the injury of the skin. The platelet gets aggregated and thereby stimulating the cascade of events like production of thrombin. The thrombin aids in the conversion of fibrinogen to fibrin and the fibrin along with the platelets forms the blood clot in order to control and stop the hemorrhage (Wilkinson, H. N et al., 2020).

Inflammatory phase:

The inflammatory phase is considered the over-lapping phase of the hemostatic phase. This phase may last for 24 hours to 2 weeks. It is characterized by vasoconstriction, secretion of pro-inflammatory agents, neutrophils and the aggregation of platelets for the induction of phagocytosis (Maheswary, T et al., 2021).

Proliferative phase:

The proliferative phase is initiated at the end of the inflammatory phase. The proliferation of fibroblast aids in the deposition of collagen and the contraction of wounds, which ultimately results in the reduction of wound size. The epithelial cells start to arrange in the normal pattern on the wounded area and followed by the angiogenesis and re-establishment of cellular network takes place. This phases ends with the formation of granulation tissue which promotes the closure of wounds (Sinno, H et al., 2013).

Remodeling phase:

The remodeling phase may take days to several months. The remodeling of dermis, epidermis, nerves, and the myofibres found in the skeletal muscle happens in the formation of functional tissue. The granulation tissue gets replaced by the collagen and so it gets converted to the scar with low tensile strength (Cañedo-Dorantes, L et al., 2019)

ROLE OF PLANTS IN WOUND HEALING:

The traditional system of healing wounds, majorly employs the natural resources and now-a- days it has gained broad range of attention among the world-wide population (Aleksandra Shedoeva et al., 2019). The Plants has an essential role in healing of various wounds from the ancient times. Various plants derived extracts have been employed in folk medicine for curing the distinct types of epithelium and skin wounds. These herbal products stimulate the rapid wound healing by accelerating the blood clotting mechanism and preventing infections (Dhanalekshmi, U et al.,

2010). The plants are regarded as the most promising agent for wound healing as it repairs the impaired skin in the natural way (Kumarasamyraja, D et al., 2012). The phytochemicals or secondary metabolites present in the plants aids in potent therapeutic activity. The medicinal plants serve as the natural repository of novel compounds with numerous biological activities (Nagori, B. P et al., 2011). It is estimated that about 73% of currently using pharmaceutical products are

derived from the natural sources like plants (Aleksandra Shedoeva et al., 2019). The utilization of plant-based medicines results in lesser side-effects and also it is cost-effective compared to other sources of medicines. Numerous scientific studies have evidenced the wound healing activity of various plant extracts along with the antimicrobial activity (Bahramsoltani et al., 2014). Some of the medicinal plants in Western Ghats used for healing wounds are listed below.

Table 1: List of some medicinal plants in Western Ghats for healing wounds

NAME OF THE PLANT	FAMILY	COMMON NAMES	NATURAL HABITAT
<i>Aegle marmelos</i>	Rutaceae	Bael, bengal quince, Japanese bitter orange, stone apple	India, Bangladesh, Sri Lanka, Nepal, Thailand, and Malaysia
<i>Aerva lanata</i>	Amaranthaceae	Knotgrass	India, South Africa, Madagascar, Saudi Arabia
<i>Allium sativum</i>	Amaryllidaceae	Garlic	Central Asia, Siberia , Northeastern Iran
<i>Aloe barbadensis Miller</i>	Asphodelaceae	Aloe vera, burn plant, medicinal aloe	Barbados, Trinidad, Mexico, Nigeria, India and Tobago
<i>Anethum graveolens</i>	Apiaceae	Dill	India, Malaysian archipelago and Japan.
<i>Azadirachta indica</i>	Meliaceae	Neem, nimtree, Indian lilac	India, Nepal, Pakistan, Bangladesh, Sri Lanka, and Maldives
<i>Calotropis gigantea</i>	Apocynaceae	Milkweed, swallow-wort	India, Cambodia, Vietnam, Bangladesh, Indonesia, Malaysia, the Philippines, Thailand, Sri Lanka, China, Pakistan, Nepal, tropical Africa

<i>Catharanthus roseus</i>	Apocynaceae	Madagascar periwinkle,graveyard plant	India, Madagascar
<i>Curcuma longa</i>	Zingiberaceae	Turmeric	Asia, India, China

HEALING PROPERTIES OF PLANTS

The wound healing activity of the plants mainly relies on pharmacological activities like anti-microbial, antioxidant, and anti-inflammatory. Then the phytoconstituents such as alkaloids, flavonoids, tannins, terpenoids, saponins, essential oils, and carbohydrates contribute to the synthesis of collagen, angiogenesis, cellular proliferation, and re-epithelialization and remodeling tissues. This ultimately results in the contraction and healing of wounds (Derakhshanfar et al.,2019).

So, the phytochemicals aid the healing of wounds via anti-microbial, antioxidant, and anti-inflammatory properties. Some of the plant extracts and their bioactive compounds act as potent free radical scavengers, which ultimately results in enhanced wound healing (Shetty et al., 2008; Geethalakshmi et al., 2013).

Table 2: Wound healing activity of selected medicinal plants

NAME OF PLANT	PARTS	EXTRACT	METHOD	CONCENTRATION	% OF CONTRACTION AND DAYS	REFERENCE
<i>Aegle marmelos</i>	Leaves	Methanol	Excision wound in male albino Swiss mice	Topical application in the	99.93±0.05 % (12 th)	Arunachalam et al., 2012
<i>Aerva lanata</i>	Leaves	Ethanol	Excision wound, Incision wound, and Dead space	300mg/kg	97.24±0.01 % (14 th day)	Devi et al., 2009
<i>Azadirachta indica</i>	Leaves	Ethanol	Linear incision wound in Albino rabbits			Munir et al., 2021
<i>Allium sativum</i>	Bulb	Ethanol	Excision wound model in swiss albino mice model	10%(w/w)	100% (16 th day)	Zuber et al., 2013
<i>Aloe barbadensis Miller</i>	Gel	Aqueous	Excision wound infemale Sprague-Dawley rats	96.4%	99.19% (16 th day)	Yadav et al., 2012
<i>Anethum graveolens</i>	Aerial parts	Essential Oil	MRSA-induced wound in BALB/c mice model	4%	100% (16 th day)	Manzuoerh et al., 2019
<i>Azadirachta indica</i>	Leaves	Ethanol	Excision model in Albino and Wister mice model	5%w/w	88.65% (15 th day)	Babu et al., 2016
<i>Calotropis gigantea</i>	Root bark	Ethanol	Excision wound	50mg /day	98.96±0.13 % (20 th day)	Deshmukh et al., 2009
<i>Catharanthus roseus</i>	Flower	Aqueous	Excision wound model in albino rats		91.27±3.45 % (18 th day)	Satish et al., 2021
<i>Curcuma longa</i>	Rhizomes	Ethanol	Excision wound in male wistar rats model	Topical application	100%(21 st day)	Charde et al., 2010

Anti- microbial effects:

The anti-microbial activities are also largely influencing the process of wound healing. The phytochemicals present in the extracts with anti-microbial effects help in managing the infections caused by various microbes in acute and chronic wounds (Rex et al., 2018). Usually, the wounds are vulnerable to infections and so the existence of micro-organisms delays the process of healing. So, the anti-microbial substances restrains the infections and also promotes rapid wound healing (Vittorazzi et al., 2016). A low rate of infections in the wounds leads to the production of numerous neutrophils, which fight against the microbes and also helps in averting the desiccation of tissues as well as necrosis (de la Secreción, 2012).

Table 3: Anti-microbial properties of selected medicinal plants

PLANT	PARTS	EXTRACTS	CONCENTRATION	TEST ORGANISMS	ZONE OF INHIBITION/ MIC	REFERENCE
<i>Aegle marmelos</i>	Fruits	Ethanol	200 µg/ml	<i>S. epidermis</i>	12.6±0.35mm	Gupta et al., 2018
				<i>S.aureus</i>	10.3±0.41mm	
				<i>E. coli</i>	14.6±0.52mm	
				<i>P.aeruginosa</i>	16.1±0.86mm	
				<i>C.albicans</i>	8.6±0.24mm	
<i>Aerva lanata</i>	Leaves	Ethanol	900 µg/ml	<i>E. coli</i>	14.05mm	Kairamkonda et al., 2017
				<i>P.vulgaris</i>	8.04mm	
				<i>B.subtilis</i>	12.08mm	
				<i>B.pumilus</i>	4.28mm	
				<i>C.albicans</i>	9.56mm	
				<i>F.oxyparum</i>	13.56mm	
				<i>C.falcatum</i>	13.76mm	
<i>Allium sativum</i>	Clove	Aqueous	100µl	<i>B. cereus</i>	21.7±1.2mm	Phan et al., 2019
				<i>L. monocytogenes</i>	33.9±1.2mm	
				<i>S. aureus</i>	19.1±1.1mm	
				<i>E.coli</i>	15.8±0.7mm	
				<i>R.mucilaginoso</i>	27.1±0.4mm	
				<i>C. albicans</i>	23.5±3.7mm	
<i>Aloe barbadensis Miller</i>	Leaves	Methanol	100mg/ml	<i>S. marcescens</i>	11.00±1.00m m	Dharajiya et al., 2017
				<i>B. cereus</i>	12.33±0.57m	
				<i>P.aeruginosa</i>	08.83±0.76m	
				<i>E.coli</i>	10.33±0.57m	
				<i>A.oryzae</i>	08.6±0.57mm	
	Sap	Acetone	6.25µg/ml	<i>E. coli</i>	31mm	Abakar et al.,
				<i>P.aeruginosa</i>	31mm	

				<i>B.subtilis</i>	30mm	2017
				<i>S.aureus</i>	30mm	
				<i>C.albicans</i>	20mm	
				<i>A.niger</i>	18mm	
<i>Anethum</i>	Dried	Essential oil	10 µL	<i>E.faecalis</i>	8.7±0.5mm	Tanruean <i>et</i>
				<i>K. pneumonia</i>	11.3±0.5mm	
<i>Azadiracht aindica</i>	Leaves	Chloroform	8mg/ml	<i>Salmonella sp</i>	8.2mm	Akoh <i>et al.</i> ,
				<i>P.mirabilis</i>	2.2mm	
				<i>S.aureus</i>	6.0mm	
				<i>S.pyogenes</i>	4.0mm	
	Twigs	Methanol	500 mg/ml	<i>S.mutans</i>	20mm	Adyanthaya <i>et al.</i> , 2014
				<i>S.salivarius</i>	10mm	
				<i>S.mitis</i>	16mm	
				<i>Lactobacillus sp</i>	22mm	
				<i>P.intermedia</i>	20mm	
				<i>C.albicans</i>	10mm	
<i>Calotropis gigantea</i>	Latex	Aqueous	100µl	<i>S.aureus</i>	30±1mm	Kumar <i>et al.</i> ,
				<i>Bacillus cereus</i>	16±1mm	
				<i>E. coli</i>	24.6±0.5mm	
				<i>Candida krusei</i>	13.6±0.5mm	
<i>Catharant hus roseus</i>	Leaves	Ethanol	100mg/ml	<i>E. coli</i>	11mm	Khalil <i>et al.</i> ,
				<i>S. aureus</i>	15mm	
				<i>C.albicans</i>	12mm	
<i>Curcuma longa</i>	rhizomes	Methanol	1mg/ml	<i>B.subtilis</i>	14.00±0.70m	Ikpeama <i>et</i> <i>al.</i> , 2015
				<i>P.aeriginosa</i>	15.00±0.80m	

				<i>S.aureus</i>	7.00±1.41mm	
				<i>E. coli</i>	9.00±0.14mm	
				<i>S.faecalis</i>	12.00±0.14mm	
				<i>Micrococcus lutes</i>	13.00±0.70mm	
				<i>A.niger</i>	20.00±0.71mm	
				<i>P.citrinin</i>	16.00±0.90mm	

Antioxidant effects:

The antioxidant properties play a vital role in the wound healing effects. The plant extracts or plant derived compounds with an increased level of antioxidants has exhibited significant wound healing activity (Lordani et al., 2018). Reactive oxygen species (ROS) are produced against the invading bacteria at the injured site. But, the existence of an increased number of ROS will affect the process of wound healing and so the plant antioxidants help in reducing the excess ROS and the adverse effects. Hence, the plant extracts and bioactive compounds with remarkable radical scavenging activity will be a potent source for wound healing (Süntar et al., 2012). The antioxidant capabilities of medicinal plants in traditional medicine for wound healing were listed below.

Table 4: Antioxidant activities of selected medicinal plants

PLANT	PARTS	EXTRACT	CONCENTRATION	METHODS	RESULTS IC50 / % OF INHIBITION	REFERENCE
<i>Aegle marmelos</i>	Leaves	Aqueous	100µl	DPPH scavenging	132.1µg/ml	Reshma and Brindha, 2014
			200µl	Reducing power activity	17.84 µg/ml	
				Superoxide anion scavenging activity	24.89 µg/ml	
<i>Aerva lanata</i>	Whole plant	Ethanol	75µg/ml	DPPH method	59.40±0.93%	Krishnamoorthi, 2015
<i>Allium</i>				DPPH		

<i>sativum</i>	Bulb	Chopped	1ml	method	21.69±3.22%	Queiroz <i>et al.</i> ,
<i>Aloe barbadensis</i>	Gel	Ethanol	400µg/ml	DPPH method	51.09±0.338 %	Gorsi <i>et al.</i> ,
<i>Anethum graveolens</i>	Leaves	Aqueous	1mg/ml	DPPH method	86%	Oshaghi <i>et al.</i> , 2016
				Superoxide scavenging activity	96%	
				Hydrogen peroxide Scavenging activity	96%	
<i>Azadirachta indica</i>	Leaves	Methanol	500 µg	DPPH method	71.23%	Pokhrel <i>et al.</i> , 2015
<i>Calotropis gigantea</i>	Flowers	Chloroform	1mg/ml	DPPH method	89.27±0.01µg/ml	Rajamohan <i>et al.</i> , 2014
<i>Catharanthus roseus</i>	Leaves	Methanol	800µg	DPPH method	81.70%	Patharajan and BalaAbirami, 2014
<i>Curcuma</i>	Rhizom	Juice	200 µl	DPPH	64.6 ± 2.4%	Maizura <i>et</i>

Anti-inflammatory effects:

The plant extracts with anti-inflammatory agents are also considered to be effective in the wound healing process. So, the phytochemicals or the secondary metabolites can act as a modulator to reduce inflammation and encourages the epithelization of impaired skin (Shukla i 2019). The inflammation may also affect the rate of wound healing. The phyto-compounds have the tendency to diminish the inflammation and its related conditions via modulating the inflammatory conditions (Ghuman et al., 2019). The anti-inflammatory properties of selected medicinal plants are enlisted below.

Table 5: Anti-inflammatory activities of selected medicinal plants

PLANT	PARTS	EXTRACT	CONCENTRATION	METHODS	RESULTS	REFERENCE
<i>Aegle marmelos</i>	Leaves	Aqueous	100 μ l	Inhibition of protein denaturation	95.64 μ g/ml	Reshma and Brindha, 2014
				Membrane stabilization activity	405.6 μ g/ml	
				Inhibition of proteinase activity	75.45 μ g/ml	
<i>Aerva lanata</i>	Whole plant	Aqueous	800 μ g/ml	Membrane stabilization method	104.4%	KV et al., 2021
				Protein denaturation method	129.3%	
<i>Allium sativum</i>	Bulb	Methanol	20mg/kg	Carrageenan paw edema in male wistar rat model	90.9%	Nithya, 2021
<i>Aloe barbadensis Miller</i>	Leaves	Aqueous	100 mg/kg	Formalin Induced Hind Paw Oedema in wistar rats	41.77%	Egesie et al., 2011

Purushothaman.T et.al.,/IJNS. Vol 1 (1): 2024,PP: 1-10				Acetic Acid-Induced Abdominal Writhing Test	68.06%	ISSN: 2584-2846
<i>Anethum graveolens</i>	Leaves	Essential oil	45.0mg/mL	In-vitro model of lipopolysaccharide (LPS)-stimulated	82.0±0.0%	Kazemi et al., 2015
<i>Azadirachta indica</i>	Bark	Ethanol	500mg/kg	Carrageenan induced rat paw inflammation	22.22±0.30%	Emran et al., 2015
<i>Calotropis gigantea</i>	Leaves	Ethanol	200mg/kg	Inhibition of albumin denaturation technique	78.57%	Va et al., 2010
<i>Catharanthus roseus</i>	Whole plant	Ethanol	300mg/ml	Membrane stabilization method	82.5±0.05%	Vinay et al., 2014
				Albumin denaturation method	83.31±0.06%	
				Heat induced hemolysis	88.82±0.06%	
<i>Curcuma Longa</i>	Leaves	Essential oil	4mg/ml	DPPH Radical Scavenging Assay	65%	Priya et al., 2012
			3.3mg/ml	ABTS Radical scavenging assay	85%	
<i>Linum Usitatissimum</i>	Seeds	Ethanol	500mg/ml	DPPH Radical Scavenging Assay	82.53±0.17954%	Amin & Thakur, 2014
				Scavenging of Hydrogen Peroxide	86.61±0.02404%	

CONCLUSION: The present review has highlighted the wound healing potency of some of the medicinal plants found in the Western Ghats. The wound healing properties of medicinal plants are influenced by their anti-microbial, anti-inflammatory as well as antioxidant activities. Therefore, many reports are also enlisted in this review for demonstrating the anti-microbial, anti-inflammatory as well as anti-oxidant efficacy of the

selected medicinal plants. So, the above-listed evidence indicated that the selected medicinal plants possess significant wound healing properties and it also promotes the healing mechanism via a natural process. Hence, the isolation of bioactive compounds from these

selected medicinal plants will benefit the development of novel medicines. But, there are still many unexplored medicinal plants, which are used by the indigenous people for wound healing. So, extensive research should be carried out to identify and validate those plants for their therapeutic benefits.

REFERENCE:

1. Abakar, H. O. M., Bakhiet, S. E., &Abadi, R. S. M. (2017). Antimicrobial activity and minimum inhibitory concentration of Aloe vera sap and leaves using different extracts. *Journal of Pharmacognosy and Phytochemistry*, 6(3), 298-303.
2. Adyanthaya, S., Pai, V., & Jose, M. (2014). Antimicrobial potential of the extracts of the twigs of *Azadirachta indica* (Neem): an in vitro study. *Journal of Medicinal Plants Studies*, 2(6), 53-57.
3. Akoh, O. U., Mac-Kalunta, O. M., &Amadi, O. K. (2021). Antimicrobial Screening, Vitamin Assay and GC MS Analysis of Chloroform Extract of *Azadirachta indica* (Neem) Leave. *Journal of Chemical Society of Nigeria*, 46(1), 0101 – 0109
4. Alachaher, F. Z., Dali, S., Dida, N., &Krouf, D. (2018). Comparison of phytochemical and antioxidant properties of extracts from flaxseed (*Linum usitatissimum*) using different solvents. *International Food Research Journal*, 25(1):75 – 82.
5. Alam, G., Singh, M. P., & Singh, A. (2011). Wound healing potential of some medicinal plants. *International Journal of Pharmaceutical Sciences Review and Research*, 9(1), 136-145.
6. Aleksandra Shedoeva, David Leavesley, Zee Upton, Chen Fan, "Wound Healing and the Use of Medicinal Plants", *Evidence-Based Complementary and Alternative Medicine*, vol. 2019, Article ID 2684108, 30 pages, 2019. <https://doi.org/10.1155/2019/2684108>
7. Amin, T., & Thakur, M. (2014). A comparative study on proximate composition, phytochemical screening, antioxidant and antimicrobial activities of *Linum usitatissimum* L.(flaxseeds). *Int. J. Curr. Microbiol. App. Sci*, 3(4), 465-481.
8. Arunachalam, K. D., Subhashini, S., &Annamalai, S. K. (2012). Wound healing and antigenotoxic activities of *Aeglemarmelos* with relation to its antioxidant properties. *J Pharm Res*, 5(3), 1492-1502.
9. Babu, K. S., Naik, V. K. M., Latha, J., &Prabhakar, V. (2016). Wound healing activity of ethanolic extract of natural products (*Azadirachta indica* bark) in albino wister rats. *World Journal of Pharmacy and Pharmaceutical Sciences*, 5(6), 1624-1632. DOI: 10.20959/wjpps20166-6949
10. Bahramsoltani, R., Farzaei, M. H., &Rahimi, R. (2014). Medicinal plants and their natural components as future drugs for the treatment of burn wounds: an integrative review. *Archives of dermatological research*, 306(7), 601-617.DOI 10.1007/s00403-014-1474-6
11. Beroual, K., Agabou, A., Abdeldjelil, M. C., Boutaghane, N., Haouam, S., &Hamdi-Pacha, Y. (2017). Evaluation of crude flaxseed (*Linumusitatissimum* L.) oil in burn wound healing in New Zealand rabbits. *African Journal of Traditional, Complementary and Alternative Medicines*, 14(3), 280-286. doi:10.21010/ajtcam.v14i3.29
12. Cañedo-Dorantes, L., & Cañedo-Ayala, M. (2019). Skin acute wound healing: a comprehensive review. *International journal of inflammation*, 2019.<https://doi.org/10.1155/2019/3706315>
13. Charde, R. M., Dhongade, H. J., Charde, M. S., & Joshi, S. B. (2010). Evaluation of Wound Healing, Anti- Inflammatory and Antioxidant Activity of Rhizomes of *Curcuma longa*. *Research Journal of Pharmacology and Pharmacodynamics*, 2(1), 42-47.
14. Dar, R. A., Shahnawaz, M., & Qazi, P. H. (2017). General overview of medicinal plants: A review. *The Journal of Phytopharmacology*, 6(6), 349-351.
15. de la Secreción, P. (2012). Assessment of antimicrobial activity and healing potential of mucous secretion of *Achatinafulica*. *Int. j. morphol*, 30(2), 365-373.
16. Deshmukh, P. T., Fernandes, J., Atul, A., &Toppo, E. (2009). Wound healing activity of *Calotropisgigantea* root bark in rats. *Journal of ethnopharmacology*, 125(1), 178-181. <https://doi.org/10.1016/j.jep.2009.06.007>
17. Devi, P., Merlin, N. J., Madhumitha, B., &Meera, R. (2009). Wound healing property of *Aervalanata* leaves extract. *Research Journal of Pharmacy and Technology*, 2(1), 210-211.

18. Dharajiya, D., Pagi, N., Jasani, H., & Patel, P. (2017). Antimicrobial activity and phytochemical screening of Aloe vera (*Aloe barbadensis* Miller). *International Journal of Current Microbiology and Applied Sciences*, 6(3), 2152-62. <https://doi.org/10.20546/ijcmas.2017.603.246>
19. Derakhshanfar, A., Moayedi, J., Derakhshanfar, G., & Fard, A. P. (2019). The role of Iranian medicinal plants in experimental surgical skin wound healing: An integrative review. *Iranian journal of basic medical sciences*, 22(6), 590–600. doi: 10.22038/ijbms.2019.32963.7873
20. Dhanalekshmi, U. M., Poovi, G., Kishore, N. M. D., Raja, M. D., & Reddy, P. N. (2010). Evaluation of wound healing potential and antimicrobial activity of ethanolic extract of *Evolvulus salsinoides*. *Annals of Biological Research*, 1(2), 49-61.
21. Egesie, U. G., Chima, K. E., & Galam, N. Z. (2011). Anti-inflammatory and analgesic effects of aqueous extract of Aloe Vera (*Aloe barbadensis*) in rats. *African Journal of Biomedical Research*, 14(3), 209-212.
22. Emran, T. B., Nasir Uddin, M. M., Rahman, A., Uddin, Z., & Islam, M. (2015). Phytochemical, antimicrobial, cytotoxic, analgesic and anti-inflammatory properties of *Azadirachta indica*: A therapeutic study. *J Bioanal Biomed S*, 12, 2-9. <http://dx.doi.org/10.4172/1948-593X.S12-007>
23. Ghuman, S., Ncube, B., Finnie, J. F., McGaw, L. J., Njoya, E. M., Cooposamy, R. M., & Van Staden, J. (2019). Antioxidant, anti-inflammatory and wound healing properties of medicinal plant extracts used to treat wounds and dermatological disorders. *South African Journal of Botany*, 126, 232-240. doi:10.1016/j.jtv.2019.09.002
24. Geethalakshmi, R., Sakravarthi, C., Kritika, T., Arul Kirubakaran, M., & Sarada, D. V. L. (2013). Evaluation of antioxidant and wound healing potentials of *Sphaeranthus amaranthoides* Burm. f. *BioMed research international*, Article ID 607109. <https://doi.org/10.1155/2013/607109>
25. Gorski, F. I., Kausar, T., & Murtaza, M. A. (2019). 27. Evaluation of antibacterial and antioxidant activity of Aloe vera (*Aloe barbadensis* Miller) gel powder using different solvents. *Pure and Applied Biology (PAB)*, 8(2), 1265-1270. <http://dx.doi.org/10.19045/bspab.2019.80068>
26. Gupta, A., Thomas, T., & Khan, S. (2018). Physicochemical, phytochemical screening and antimicrobial activity of *Aegle marmelos*. *Pharmaceutical and Biosciences Journal*, 6(3), 17-24. DOI: 10.20510/ukjpb/6/i3/173548
27. Han, G., & Ceilley, R. (2017). Chronic wound healing: a review of current management and treatments. *Advances in therapy*, 34(3), 599-610.
28. Ikpeama, A., Onwuka, G. I., & Nwankwo, C. (2014). Nutritional composition of Tumeric (*Curcuma longa*) and its antimicrobial properties. *International Journal of Scientific and Engineering Research*, 5(10), 1085-1089.
29. Jeyaprakash, K., Ayyanar, M., Geetha, K. N., & Sekar, T. (2011). Traditional uses of medicinal plants among the tribal people in Theni District (Western Ghats), Southern India. *Asian Pacific Journal of Tropical Biomedicine*, 1(1), S20-S25.
30. Kairamkonda, M., Godishala, V., & Kandi, V. (2017). Identification of Newer Antimicrobial Agents: A Study of In Vitro Antibacterial and Antifungal Activities of Leaf extracts of Medicinal Plant *Aervalanata* (L.) Juss. exSchult. *American Journal of Microbiological Research*, 5(3), 66-70. DOI:10.12691/ajmr-5-3-3
31. Kazemi, M. (2015). Chemical composition and antimicrobial, antioxidant activities and anti-inflammatory potential of *Achillea millefolium* L., *Anethum graveolens* L., and *Carum copticum* L. essential oils. *Journal of Herbal Medicine*, 5(4), 217-222. <https://doi.org/10.1016/j.hermed.2015.09.001>
32. Khalil, A. (2012). Antimicrobial activity of ethanol leaf extracts of *Catharanthus roseus* from Saudi Arabia. In *2nd International Conference on Environment Science and Biotechnology* (Vol. 48, No. 2, pp. 6-11). DOI: 10.7763/IPCBE
33. Krishnamoorthi, R. (2015). Phytochemical analysis and antioxidant property of *Aervalanata*. *International journal of pharmacognosy*, 2(8), 426-429.
34. Kumar, G., Karthik, L., & Rao, K. B. (2010). Antimicrobial activity of latex of *Calotropis gigantea* against pathogenic microorganisms-an in vitro study. *Pharmacologyonline*, 3(3), 155-163.
35. Kumarasamyraja, D., Jeganathan, N. S., & Manavalan, R. (2012). A review on medicinal plants with potential wound healing activity. *Int J Pharm PharmSci*, 2, 105-11.
36. KV, S. K., Likitha, E., Yaswantha, B., Umakavya, C., Sri, K. U., Suneel, M., ... & Vamsi, V. (2021). Evaluation of Anti-Inflammatory Activity on Whole Plant of *Aervalanata* (L.) by In-Vitro Methods. *Future Journal of Pharmaceuticals and Health Sciences*, 1(3), 56-61. DOI: <https://doi.org/10.26452/>
37. Lordani, T. V. A., de Lara, C. E., Ferreira, F. B. P., de Souza Terron Monich, M., Mesquita da Silva, C., Felicetti Lordani, C. R., ... & Lonardon, M. V. C. (2018). Therapeutic effects of medicinal plants on cutaneous wound healing in humans: a systematic review. *Mediators of inflammation*, 2018. Article ID 7354250 | <https://doi.org/10.1155/2018/7354250>
38. Maheswary, T., Nurul, A. A., & Fauzi, M. B. (2021). The Insights of Microbes' Roles in Wound Healing: A Comprehensive Review. *Pharmaceutics*, 13(7), 981-1013. <https://doi.org/10.3390/pharmaceutics13070981>

39. Maizura, M., Aminah, A., & Wan Aida, W. M. (2011). Total phenolic content and antioxidant activity of kesum (*Polygonum minus*), ginger (*Zingiberofficinale*) and turmeric (*Curcuma longa*) extract. *International Food Research Journal*, 18(2):526-531.
40. Manzuoerh, R., Farahpour, M. R., Oryan, A., & Sonboli, A. (2019). Effectiveness of topical administration of *Anethumgraveolens* essential oil on MRSA-infected wounds. *Biomedicine & Pharmacotherapy*, 109, 1650-1658. <https://doi.org/10.1016/j.biopha.2018.10.117>
41. Munir, M., Shah, S. N. H., Almas, U., Khan, F. A., Zaidi, A., Bukhari, S. M., & Murtaza, G. (2021). An assessment of the wound healing potential of a herbal gel containing an *Azadirachta indica* leaf extract. *Veterinární medicína*, 66(3), 99-109. <https://doi.org/10.17221/46/2020-VETMED>
42. Nagori, B. P., & Solanki, R. (2011). Role of medicinal plants in wound healing. *Research Journal of Medicinal Plant*, 5(4), 392-405.
43. Narender, B. R., Tejaswini, S., Sarika, M., Karuna, N., Shirisha, R., & Priyanka, S. (2016). Antibacterial and antifungal activities of *Linum usitatissimum* (Flax seeds). *Int J Pharm Educ Res*, 3, 4-8.
44. Nithya, V. (2021). Anti-Inflammatory Activity of *Allium sativum* Linn., in Wistar Albino Rats. *Inventi Rapid: Ethnopharmacology Vol. 2, Issue 1:1-2*.
45. Oshaghi, E. A., Khodadadi, I., Tavilani, H., & Goodarzi, M. T. (2016). Aqueous extract of *Anethum Graveolens* L. has potential antioxidant and antiglycation effects. *Iranian Journal of Medical Sciences*, 41(4), 328-333.
46. Patharajan, S., & Bala Abirami, S. (2014). Antioxidant activity and phytochemical analysis of fractionated leaf extracts of *Catharanthus roseus*. *Int J Pharm*, 1(2), 138-143.
47. Phan, A. D. T., Netzel, G., Chhim, P., Netzel, M. E., & Sultanbawa, Y. (2019). Phytochemical characteristics and antimicrobial activity of Australian grown garlic (*Allium sativum* L.) cultivars. *Foods*, 8(9), 358. doi:10.3390/foods8090358
48. Pokhrel, B., Rijal, S., Raut, S., & Pandeya, A. (2015). Investigations of antioxidant and antibacterial activity of leaf extracts of *Azadirachta indica*. *African Journal of Biotechnology*, 14(46), 3159-3163. DOI: 10.5897/AJB2015.14811
49. Priya, R., Prathapan, A., Raghu, K. G., & Menon, A. N. (2012). Chemical composition and in vitro antioxidative potential of essential oil isolated from *Curcuma longa* L. leaves. *Asian Pacific Journal of Tropical Biomedicine*, 2(2), S695-S699.
50. Queiroz, Y. S., Ishimoto, E. Y., Bastos, D. H., Sampaio, G. R., & Torres, E. A. (2009). Garlic (*Allium sativum* L.) and ready-to-eat garlic products: in vitro antioxidant activity. *Food chemistry*, 115(1), 371-374.
51. Rajamohan, S., Kalaivanan, P., Sivangnanam, H., & Rajamanickam, M. (2014). Antioxidant, Antimicrobial activities and GC-MS analysis of *Calotropis gigantea* white flowers. *J. Phytopharmacol*, 3, 405-409.
52. Rawat, S., Singh, R., Thakur, P., Kaur, S., & Semwal, A. (2012). Wound healing agents from medicinal plants: a review. *Asian Pacific Journal of Tropical Biomedicine*, 2(3), S1910-S1917. [https://doi.org/10.1016/S2221-1691\(12\)60520-6](https://doi.org/10.1016/S2221-1691(12)60520-6)
53. Reshma, A. K., & Brindha, P. (2014). In vitro anti-inflammatory, antioxidant and nephroprotective studies on leaves of *Aegle marmelos* and *Ocimum sanctum*. *Asian J Pharm Clin Res*, 7(4):121-129.
54. Rex, J. R. S., Muthukumar, N. M. S. A., & Selvakumar, P. M. (2018). Phytochemicals as a potential source for anti-microbial, antioxidant and wound healing-a review. *MOJ Biorg Org Chem*, 2(2), 61-70.
55. Sharma, P., Manchanda, R., Goswami, R., & Chawla, S. (2020). Biodiversity and therapeutic potential of medicinal plants. In *Environmental Concerns and Sustainable Development* (pp. 27-44). Springer, Singapore. https://doi.org/10.1007/978-981-13-6358-0_2
56. Shetty, S., Udupa, S., & Udupa, L. (2008). Evaluation of antioxidant and wound healing effects of alcoholic and aqueous extract of *Ocimum sanctum* Linn in rats. *Evidence-Based Complementary and Alternative Medicine*, 5(1), 95-101. doi:10.1093/ecam/nem004
57. Shukla, S. K., Sharma, A. K., Gupta, V., & Yashavardhan, M. H. (2019). Pharmacological control of inflammation in wound healing. *Journal of tissue viability*, 28(4), 218-222. <https://doi.org/10.1016/j.jtv.2019.09.002>
58. Sinno, H., & Prakash, S. (2013). Complements and the wound healing cascade: an updated review. *Plastic Surgery International Volume 2013, Article ID 146764, 7 pages* <http://dx.doi.org/10.1155/2013/146764>
59. Süntar, I., Akkol, E. K., Nahar, L., & Sarker, S. D. (2012). Wound healing and antioxidant properties: do they coexist in plants?. *Free Radicals and Antioxidants*, 2(2), 1-7. DOI: 10.5530/ax.2012.2.1
60. Tanruean, K., Kaewnarin, K., & Rakariyatham, N. (2014). Antibacterial and antioxidant activities of *Anethumgraveolens* L. dried fruit extracts. *Chiang Mai Journal of Science*, 41(3), 649-660.
61. VA, J., Usman, M. R. M., Salunkhe, P. S., & Gagrani, M. B. (2010). Anti-inflammatory Activity of *Calotropis gigantea* Linn. Leaves Extract on In-vitro Models. *IJCP Review and Research*, 1(2), 1-5.
62. Vinay Gupta, Archana Prakash, and Abhishek Mathur. (2014). Screening of solvent extracts of *Catharanthus roseus* for the isolation and anti-inflammatory compound. *Journal of drug discovery and therapeutics*, 2, 20:2014, 78-88.
63. Vittorazzi, C., Endringer, D. C., Andrade, T. U. D., Scherer, R., & Fronza, M. (2016). Antioxidant, antimicrobial and wound healing properties of *Struthanthus vulgaris*. *Pharmaceutical biology*, 54(2), 331-337. DOI: 10.3109/13880209.2015.1040515

64. Wilkinson, H. N., & Hardman, M. J. (2020). Wound healing: Cellular mechanisms and pathological outcomes. *Open biology*, 10(9), 200223.
65. Yadav, K. H., Kumar, J. R., Basha, S. I., Deshmukh, G. R., Gujjula, R., & Santhamma, B. (2012). Wound healing activity of topical application of Aloe vera gel in experimental animal models. *International Journal of Pharma and Bio Sciences*, 3(2), 63-72.
66. Zuber, M., Rajesh, V., Anusha, K., Reddy, C. R., & Tirupathi, A. (2013). Wound healing activity of ethanolic extract of *Allium sativum* on alloxan induced diabetic rats family (Liliaceae). *International Journal of Science Inventions Today*, 2(1), 40-57.
67. Banu, D. A. S., Sheela, S., Muralimohan, N., Saranya, N., Kolar, A. B., & Santhya, V. S. (2023, December 19). Importance and Benefits of Green Audits to Education Institutions and Industrial Sectors. *International Journal of Environment and Climate Change*, 13(12), 349–356. <https://doi.org/10.9734/ijecc/2023/v13i123690>
68. Kavitha, R., Sathish, S. S., Vinoliya, V., Jahirhussain, G., & Basha Kolar, A. (2023, August 24). Green synthesis, characterization and biological studies on in vitro cultured fern – *Lygodium microphyllum* (Cav.) R. Br. *Biochemical and Cellular Archives*, 23(2). <https://doi.org/10.51470/bca.2023.23.2.975>
69. Saminathan, M., Muruganandam, A., Arumugam, M., & Basha Kolar, A. (2023, September 14). Genetic diversity analysis of *Madhuca longifolia* (Koen) Macbr. populations by using RAPD markers. *Biochemical and Cellular Archives*, 23(2). <https://doi.org/10.51470/bca.2023.23.2.751>
70. Kolar, A. B., Sheik Mohamed, S., Shareef Khan, M., & Ghouse Basha, M. (2021, January 15). DIRECT PLANT REGENERATION FROM MATURE NODAL EXPLANTS OF ANDROGRAPHIS ECHIOIDES (L.) NEES – A VALUABLE MEDICINAL PLANT. *PLANT ARCHIVES*, 21(Suppliment-1), 1842–1848. <https://doi.org/10.51470/plantarchives.2021.v21.s1.296>
71. Amzad Basha Kolar*, Palanivel.S, M. Sheik Noor Mohamed, S. Sheik Mohamed, M. Shareef Khan, Aakash Raj.S.G., Mohammed Ibrahim.V and Mohammed Nowshath. A. (2021). FLORISTIC STUDY ON ANGIOSPERMS SURROUNDING THE MEDAVAKKAM LAKE, CHENGALPATTU DISTRICT, TAMIL NADU, INDIA. *Plant Archives*, 21(1), 1953–1962. <https://doi.org/10.51470/PLANTARCHIVES.2021.v21.no1.271>
72. Amzad Basha Kolar, Edwin Raj Esackb, L. Vivekanandanb, M. Ghouse Basha. (2015). K. R. Kavitha, A.K. Bopaiah and Amzad Basha Kolar. (2016). CHEMICAL COMPOSITION OF THE ESSENTIAL OIL FROM THE LEAVES OF *CIPADESSA BACCIFERA* (ROTH.) MIQ. *International Journal of Pharmaceutical Sciences and Research*, 7(1), 392–396. *The Journal of Ethnobiology and Traditional Medicine*, 936–957.